

*Keep and promote biodiversity at polluted sites under phytomanagement*

**Carlos Garbisu, Itziar Alkorta, Petra Kidd, Lur Epelde & Michel Mench**

**Environmental Science and Pollution Research**

ISSN 0944-1344  
Volume 27  
Number 36

Environ Sci Pollut Res (2020)  
27:44820-44834  
DOI 10.1007/s11356-020-10854-5

**Your article is protected by copyright and all rights are held exclusively by Springer-Verlag GmbH Germany, part of Springer Nature. This e-offprint is for personal use only and shall not be self-archived in electronic repositories. If you wish to self-archive your article, please use the accepted manuscript version for posting on your own website. You may further deposit the accepted manuscript version in any repository, provided it is only made publicly available 12 months after official publication or later and provided acknowledgement is given to the original source of publication and a link is inserted to the published article on Springer's website. The link must be accompanied by the following text: "The final publication is available at [link.springer.com](http://link.springer.com)".**



# Keep and promote biodiversity at polluted sites under phytomanagement

Carlos Garbisu<sup>1</sup> · Itziar Alkorta<sup>2</sup> · Petra Kidd<sup>3</sup> · Lur Epelde<sup>1</sup> · Michel Mench<sup>4</sup>

Received: 3 June 2020 / Accepted: 13 September 2020 / Published online: 25 September 2020  
© Springer-Verlag GmbH Germany, part of Springer Nature 2020

## Abstract

The phytomanagement concept combines a sustainable reduction of pollutant linkages at risk-assessed contaminated sites with the generation of both valuable biomass for the (bio)economy and ecosystem services. One of the potential benefits of phytomanagement is the possibility to increase biodiversity in polluted sites. However, the unique biodiversity present in some polluted sites can be severely impacted by the implementation of phytomanagement practices, even resulting in the local extinction of endemic ecotypes or species of great conservation value. Here, we highlight the importance of promoting measures to minimise the potential adverse impact of phytomanagement on biodiversity at polluted sites, as well as recommend practices to increase biodiversity at phytomanaged sites without compromising its effectiveness in terms of reduction of pollutant linkages and the generation of valuable biomass and ecosystem services.

**Keywords** Contaminated soil · Metal · Metallophytes · Phytoremediation · Trace elements

## Introduction

The notion of phytomanagement is based on the combination of (i) a sustainable reduction of pollutant linkages at degraded sites with (ii) the generation of valuable products and essential ecosystem services. In other words, its main purpose is to grow profitable plants to minimise pollutant-induced environmental risks while maximising economic and/or ecological revenues. It is often claimed that one of the potential benefits

of phytomanagement is the possibility to enhance biodiversity in the degraded site under recovery. Pertinently, it must be strongly emphasised that some polluted sites, most relevantly mining sites, can harbour a unique biodiversity that must be carefully preserved. In any event, protecting biodiversity is of the utmost importance as human well-being depends upon biodiversity in many different ways (Naeem et al. 2016). In consequence, under the current scenario of global change and biodiversity loss, it is crucial to use as many tools as possible to preserve the fabric of life and the natural capital on which our survival and well-being depend. Biodiversity is known to be critical for the supply of ecosystem services and, then, it is not surprising that much research effort has been directed at understanding how biodiversity impacts ecosystem functioning and resilience, and concomitantly the sustainable provision of goods and ecosystems services. This aspect has special relevance within the phytomanagement framework since, as described above, the main purpose of phytomanagement is to grow profitable plants in order to minimise pollutant-induced environmental risks while maximising economic and/or ecological revenues in terms of products and ecosystem services. However, when implementing actions to promote such biodiversity in phytomanaged sites, in most cases, the only initiative is to enhance the number of different plant species grown for phytomanagement purposes. We must overcome such incomplete approach by widening our understanding of how the

---

Responsible Editor: Elena Maestri

✉ Carlos Garbisu  
cgarbisu@neiker.eus

<sup>1</sup> Department of Conservation of Natural Resources, Soil Microbial Ecology Group, NEIKER-Basque Institute for Agricultural Research and Development, Basque Research and Technology Alliance (BRTA), Parque Científico y Tecnológico de Bizkaia P812, E-48160 Derio, Spain

<sup>2</sup> Department of Biochemistry and Molecular Biology, University of the Basque Country, P. O. Box 644, 48080 Bilbao, Spain

<sup>3</sup> Consejo Superior de Investigaciones Científicas (CSIC), Instituto de Investigaciones Agrobiológicas de Galicia (IIAG), 15780 Santiago de Compostela, Spain

<sup>4</sup> INRAE, BIOGECO, University of Bordeaux, F-33615 Pessac, France

different taxonomic groups can be positively or negatively affected by phytomanagement practices. In addition, the unique biodiversity present in some polluted sites can be negatively affected by the implementation of phytomanagement practices. In this review paper, the importance of promoting (i) measures to minimise the potential adverse impact of phytomanagement on biodiversity and (ii) practices to increase biodiversity at phytomanaged sites is highlighted.

## Phytomanagement: a sustainable gentle remediation option

As a result of a wide variety of anthropogenic activities and accidental spills, many soils are currently polluted with a myriad of potentially toxic compounds, such as trace elements (TEs), mineral oils, polycyclic aromatic hydrocarbons (PAHs), polychlorinated biphenyls (PCBs) and pesticides. Unfortunately, the remediation of polluted soils is often a very expensive, environmentally disruptive activity, especially at large sites and/or in those soils simultaneously polluted with several contaminants inducing adverse effects on biological receptors (Agnello et al. 2016).

Opportunely, in the last decades, various Gentle Remediation Options (GROs) have been developed as more cost-effective, environmentally friendly and aesthetically pleasing technologies for the remediation of large areas with polluted soils from mild up to medium levels of contamination (Vangronsveld et al. 2009; Kidd et al. 2015; Mench et al. 2018). Among them, phytoremediation and phytomanagement have shown their great potential, in the long term, for the sustainable remediation of polluted sites due to their capacity to combine an effective mitigation of pollutant-induced risks with the provision of valuable plant biomass and ecosystem services (Mench et al. 2018).

The term *phytoremediation* refers to a set of sustainable phytotechnologies focused on the use of plant species to remediate polluted sites, mainly those affected by the presence of TEs via the phytoextraction or phytostabilisation options, which aim at (i) decreasing the available soil TEs, through plant uptake and accumulation in the harvestable plant parts, or (ii) reducing the labile (“bioavailable”) TE pool usually by combining the growth of TE-excluding plants with the application of soil amendments (Garbisu and Alkorta 2001; Alkorta et al. 2004a, b). However, the commercial application of phytoextraction has been seriously hampered by its intrinsic limitations, e.g. the long time required to effectively extract TEs from medium and highly polluted soils, root depth, lack of plants that can accumulate more than one or two TEs, and decrease of metal(loid) market prices. In turn, one constraint for the application of phytostabilisation is that many risk-assessment regulations for soil remediation are still based on total soil TEs, not on their bioavailable concentrations or site-

specific risk assessment. Paradoxically, the harmful effects of TEs on soil biota and, hence, soil health, are related to the sensitivity/tolerance of living organism populations and the bioavailable pool rather than total metal(loid) concentrations (Kumpiene et al. 2009, 2017), the bioavailable fraction being subject to uptake by soil organisms, leaching and transfer to other environmental media (Madejón et al. 2006).

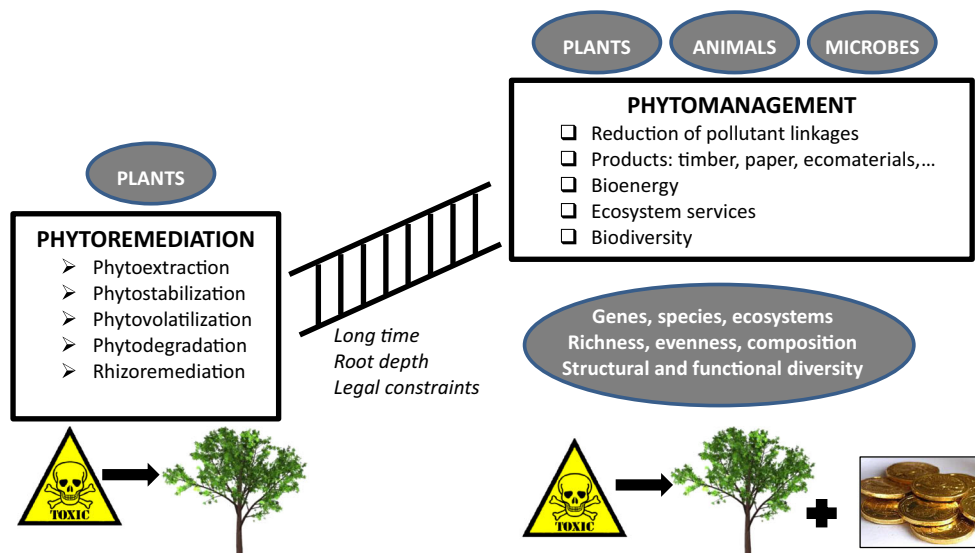
In any event, for effective clean-up of TE-polluted soils, the combination of different approaches, e.g. phytoextraction with hyperaccumulators, chelate-assisted phytoextraction, phytostabilisation, microbe-assisted phytoremediation (bioaugmentation), traditional breeding and/or genetic engineering of phytoremediation plants, appears necessary to increase the applicability of phytoremediation in the future (Yang et al. 2020a).

To overcome the phytoremediation limitations, the concept of *phytomanagement* evolved (Fig. 1), combining a sustainable reduction in pollutant linkages with the generation of plant biomass (mainly non-food crops) and ecosystem services (Mench et al. 2009; Robinson et al. 2009; Fässler et al. 2010; Robinson and McIvor 2013; Cundy et al. 2016; Burges et al. 2018). The phytomanagement objective is to grow profitable plants to control the bioavailable pool of soil pollutants (e.g. TEs), thereby minimising environmental risks, while maximising economic and ecological revenues (Vangronsveld et al. 2009). In this way, phytomanagement (commonly based on the interactions among plants, microorganisms and soil amendments) is often considered a “holding strategy” for vacant sites until their remediation is undertaken according to future land use (Mench et al. 2018). Most importantly, compared with other remediation technologies, the requirements of phytomanagement for chemicals and energy are much lower, as well as the total cost, making it a viable strategy for the remediation of large polluted areas (Thewys et al. 2010; Kuppens et al. 2015; Giagnoni et al. 2020).

The production of valuable plant biomass (for timber, bioenergy, biofortified products, ecomaterials, etc.) is considered essential for the commercial success of phytomanagement (Conesa et al. 2012). Energy crops, e.g. *Miscanthus* spp., *Ricinus communis* L. and *Brassica napus* L., can be grown for biofuel production (Burges et al. 2018). Other plant species can be grown for the production of biochar, raw materials (oil, paper, chemicals, essential oils, etc.), medicinal purposes, etc. (Pandey et al. 2016). Likewise, the growth of fast-growing trees opens the possibility to phytoextract some metals in excess (e.g. Cd, Zn, Ni, U, Cs and Sr) while producing biomass for bioenergy and products (e.g. timber, resin, adhesives). Other phytomanagement options are aimed at removing the bioavailable metal(loid) fraction, the so-called bioavailable contaminant stripping (BCS), while providing ecosystem services and feedstocks for biomass-processing (bio)technologies (Herzig et al. 2014).

Since the 2010s, an increasing attention is developing on the capacity of phytomanagement options to provide a wide variety of co-benefits and ecosystem services, such as primary

**Fig. 1** Evolution from phytoremediation to phytomanagement



production, control of soil erosion, water runoff/drainage management, carbon sequestration, amenity and recreation, aesthetic value, habitat for animals and microorganisms, and biodiversity (Evangelou et al. 2015; Kidd et al. 2015; Cundy et al. 2016; Simek et al. 2017; Touceda-González et al. 2017a; Xue et al. 2018). Strictly speaking, biodiversity per se is not an ecosystem service (Haines-Young and Potschin 2010); rather, biodiversity supports the flow of vital ecosystem services that we depend upon (in other words, biodiversity forms the biological infrastructure that supports the provision of ecosystem services). Indeed, the ecosystem services (and, concomitantly, human well-being) depend essentially on the structures and processes generated by living organisms and their interactions with, and processing of, abiotic materials (Haines-Young and Potschin 2010; IPBES 2019). Although biodiversity has intrinsic value by itself (and, then, it should be preserved in its own right), its utilitarian value has increasingly become the central focus of the debates on the need to preserve our natural capital (Chan et al. 2007; Haines-Young and Potschin 2010).

The phytomanagement capacity to promote biodiversity in polluted sites is of key importance, as we are currently at a crucial juncture in human history, with biodiversity being lost at an accelerating pace due to an increasingly affluent human population, climate change, uncontrolled development and habitat destruction (Sandifer et al. 2015; IPBES 2019). Taking into consideration the links between biodiversity, ecosystem functioning, ecosystem services and human well-being (Cardinale et al. 2006; Naeem et al. 2016), now more than ever, the importance of promoting biodiversity must be emphasised.

### Unique biodiversity at polluted sites

One of the recurrently mentioned potential benefits of phytomanagement is the possibility to increase biodiversity

in the polluted land in question. However, the unique biodiversity often present in many polluted sites (in particular, long-term abandoned mine sites) can be severely impacted by the implementation of phytomanagement practices, even resulting in the local extinction of endemic ecotypes or species of great conservation value. Here, we highlight the importance of promoting measures to minimise the potential adverse impact of phytomanagement on biodiversity at polluted sites. After all, some polluted sites, most relevantly mining sites, harbour a unique biodiversity that must be painstakingly preserved. Actually, derelict soils can provide an interesting biodiversity for a variety of uses (Vincent et al. 2018).

In particular, there is a need to conserve metallophytes (i.e. unique plant species that have evolved to survive on soils with high TE levels), which are nowadays increasingly under threat of extinction from mining activity (Whiting et al. 2004; Batty 2005; Baker et al. 2010; Paul et al. 2018). Indeed, regrettably, metalliferous ecosystems are presently threatened at a global scale by the growth of mining activities with concomitant extinction risks for metallophyte diversity (Whiting et al. 2004; Séleck et al. 2013).

Metallophytes are the consequence of powerful selective pressures over long evolutionary times as a result of the presence of high total soil TEs (Ginocchio and Baker 2004). The intensity and duration of the sustained evolutionary exposure to these high TE levels direct the degree of specialisation of the TE resistance trait. Thus, some populations of plant species can evolve TE resistance within a few years, for example, around metal smelters, if the selection pressure is high enough (Barrutia et al. 2011a). Populations of pseudometallophytes present a greater capacity to withstand phytotoxicity induced by TE excess, as compared with other populations of the same plant species from non-polluted sites (Whiting et al. 2004; Barrutia et al. 2011a). But, as the duration of TE exposure increases, the mechanisms that allow survival and growth in the presence of

high TE levels become gradually more specialised, resulting in true metallophytes or eumetallophytes that have developed evolutionary mechanisms to live and thrive on metalliferous soils. As a matter of fact, true metallophytes have often diverged genetically and morphologically to form new taxa endemic to their native metalliferous soils (Barrutia et al. 2011a). Regrettably, their restricted geographic range is, partly, responsible for the current high rates of population decline or, what is worse, irreversible extinction.

Plants growing in TE-polluted sites can be classified as (1) *excluders*: these plants limit TE uptake and translocation and, then, maintain low TE concentrations in their aerial tissues; (2) *indicators*: these plants accumulate TEs in their harvestable parts at concentrations similar to those present in the polluted soil; and (3) *accumulators/hyperaccumulators*: these plants increase TE uptake, translocation and accumulation in their aboveground biomass reaching levels that far exceed those present in the polluted soil (van der Ent et al. 2013, 2015a; Malik et al. 2017; Massoura et al. 2004; Reeves et al. 2017a). Particularly, hyperaccumulators are exceptional plants that accumulate metal(loid)s in their tissues to levels that can be hundreds or thousands of times greater than common ranges in other plant species (van der Ent et al. 2013), and whose ecology is an active field of research, focusing on anti-herbivore defences, allelopathy and biotic interactions (Reeves et al. 2017b). Logically, it is important to protect not only TE hyperaccumulators as nature's oddities but also TE excluders, indicators and accumulators.

Apart from their intrinsic value as remarkable rare species, metallophytes are suitable candidates for the revegetation of mining sites, as well as for the implementation of phytoremediation (phytoextraction/phytomining, phytostabilisation) and phytomanagement initiatives (van der Ent et al. 2015b; Rosenkranz et al. 2019; Corzo Remigio et al. 2020). Thus, TE excluders and hyperaccumulators have been extensively used for phytostabilisation and phytoextraction purposes, respectively (Hernández-Allica et al. 2006; Epelde et al. 2008, 2009, 2010; Barrutia et al. 2009; Pardo et al. 2014; Garaiurrebaso et al. 2017). There is nowadays an increasing interest in the use of native plant species and populations for the revegetation of TE-polluted sites, as opposed to non-native, introduced species (Parraga-Aguado et al. 2014; Chen et al. 2019). In this respect, a sturdy commitment to conservation of metallophyte biodiversity is self-evident (Whiting et al. 2002).

Then, before starting any phytomanagement initiative, it is imperative to study the native vegetation of the polluted site in search of potential candidates (e.g. metallophytes) for conservation purposes. If such candidates are identified, then, an area of the site (preferably, the area where the most interesting plant species have been identified) must be left unmanaged for conservation purposes (and, if needed, protection barriers must be installed).

In addition to protecting the natural environment of valued and treasured plant species (i.e. in situ conservation in biotope “islands”), efforts must also be directed at conserving them ex situ, that is to say, in germplasm banks, seed gardens, arboreta, botanic gardens, etc. (Whiting et al. 2004), ideally maintaining the required degree of contaminant exposure (otherwise, contaminant-sensitive, non-adapted individuals might again become dominant in the plant population after a few cultivations). Additionally, when designing a strategy to preserve the valuable native vegetation at the polluted site, attention should also be paid to plant assemblages (e.g. metalliferous distinctive plant communities).

Apart from the presence of unique metallophytes in TE-polluted sites, these degraded environments can also harbour a valuable microbial diversity that can likewise be used for phytoremediation and/or phytomanagement initiatives. For instance, TE-resistant plant growth-promoting rhizobacteria and endophytes, as well as TE-resistant mycorrhiza, can be isolated from TE-polluted sites and, subsequently, be used to improve plant survival, growth and performance under the harsh conditions usually present in many TE-polluted sites, particularly mining sites (Weyens et al. 2011; Ma et al. 2015; Burges et al. 2016, 2017; Harrison and Griffin 2020). For instance, rhizobacterial inoculants (e.g. *Arthrobacter nicotinovorans* SA40) have been shown to improve nickel phytoextraction by the hyperaccumulator *Alyssum pintodasilvae* (Cabello-Conejo et al. 2014). Similarly, the inoculation of ultramafic soils with *Microbacterium arabinogalactanolyticum* AY509224 increased soil Ni extractability and uptake by *Alyssum murale* (Abou-Shanab et al. 2006). The inoculation with TE-resistant plant growth-promoting bacteria has been reported to enhance the biomass of different plant species (e.g. *Brassica juncea*, *Ricinus communis*, *Helianthus annuus* and *Sedum alfredii*) growing in TE-polluted soils (Dell'Amico et al. 2008; Jiang et al. 2008; Mastretta et al. 2009; Zaidi et al. 2006). Likewise, Kolbas et al. (2015) reported the positive effects of endophytic bacteria for Cu phytoextraction by sunflower plants. Truyens et al. (2014) found that inoculation of *Agrostis capillaris* plants with endophytes can be beneficial for their establishment during phytoextraction and phytostabilisation of Cd-polluted soils. The co-inoculation of *Paenibacillus mucilaginosus* and *Sinorhizobium meliloti* in Cu-contaminated soil planted with *Medicago sativa* improved alfalfa growth and decreased Cu accumulation in shoots, compared with the uninoculated control (Ju et al. 2020). The inoculation of *Pseudomonas vancouverensis* promoted As accumulation efficiency in *Pteris vittata* and *Pteris multifida* (Yang et al. 2020b). However, the effect of bacterial inoculants on plant growth and TE accumulation has been shown to be plant species-specific (Becerra-Castro et al. 2012).

Apart from the recognised conservation value of metallophytes present at TE-polluted sites, plant species and,

interestingly, microbial (bacteria, fungi) populations living in soils polluted with organic compounds must also be considered for conservation purposes, owing to their intrinsic value as well as their potential use for the rhizoremediation (i.e. degradation of pollutants by rhizosphere bacteria) (Barrutia et al. 2011b; Lacalle et al. 2018; Brereton et al. 2020) and bioremediation of organically polluted soils (Garbisu et al. 2017; Anza et al. 2018; Megloui et al. 2019; Villaverde et al. 2019). Since bacterial and fungal strains isolated from organically polluted soils can then be used for bioremediation via bioaugmentation purposes, after thoroughly testing their degrading capabilities and potentials, it is recommended to keep them in a microbial bank for possible future biotechnological applications.

## Managing biodiversity during phytomanagement

### Phytomanagement under the current scenario of climate change

The negative consequences of climate change can nowadays be undoubtedly identified in the more frequent alteration of natural and agricultural ecosystems, owing to, for instance, higher temperatures, extreme droughts and storms, and an increased likelihood of heat waves and heavy precipitation episodes (Alkorta et al. 2017). Not surprisingly, plant survival and growth are being significantly altered under changing climatic conditions. Furthermore, increasing CO<sub>2</sub> concentrations in the atmosphere are currently changing the physiology of plants, affecting, among other aspects, their growth rate.

Specifically, regarding the choice of plant species for phytomanagement in semi-arid and arid regions (e.g. southern Europe) (Pulighe et al. 2019), and taking into account the critical importance of an adequate water regime for the success of revegetation programmes, special attention should be paid to the selection of drought-resistant plant species and ecotypes, since the duration and frequency of extreme droughts is nowadays increasing in many semi-arid and arid regions (Risueño et al. 2020).

The possibility of irrigating phytomanagement crops is decidedly controversial, since water is an increasingly scarce resource in many parts of the world. A proper sustainable management of water resources is currently one of the greatest challenges for our society worldwide. Above all, we must first ensure availability of good quality water for human consumption and agricultural purposes. Regrettably, in the coming decades, the problem of water scarcity will probably get worse than it is now. Predictably, an increase in the world human population will imply more water for human consumption and agricultural production (agriculture accounts for around 70% of the water currently used in the world). Then, it follows that

the consumption of good quality water for irrigation of phytomanaged sites is, in general, not considered a valid option, especially in semi-arid and arid regions. An alternative is the use of wastewater for irrigation. An appealing, and currently attention-grabbing, option is the possibility of treating such wastewater by means of rhizofiltration (i.e. the use of plant roots and associated microbes to absorb, concentrate and precipitate pollutants, especially TEs, from polluted effluents and waters) and/or biodegradation, notably using constructed wetlands (CWs) and floating islands (Dushenkov et al. 1995; Schröder et al. 2007; Zhang et al. 2007; Olguin et al. 2017).

Urban wastewater is known to contain nitrogen, phosphorus and other nutrients, leading to an extra beneficial effect for plant growth through fertilisation. Irrigation with wastewater is only recommended for non-food and non-fodder crops, and then, it would be an ideal option for phytomanagement. Evidently, it would be beneficial to have an efficient urban wastewater treatment plant closed to the site to be phytomanaged, so that the wastewater, directly or preferably after rhizofiltration in a CW does not need to be transported a long distance. Interestingly, CWs can also be used to treat acid mine drainage, and then, there is the possibility of reusing the treated water to eventually irrigate mine tailings (Pat-Espadas et al. 2018).

Therefore, especially in semi-arid and arid regions of the world, for phytomanagement purposes, it is recommended to select plant species that are resistant to water stress, extreme droughts and heat waves for increasing the long-term success of the phytomanagement strategy (Risueño et al. 2020). For instance, as water supply and its distribution during the crop cycle is a key limiting factor for crop production in SW France, the sunflower and tobacco ability to stand more frequent heat waves and long droughts is certainly an advantage (Kidd et al. 2015; Mench et al. 2018). Although hundreds of plant species are suitable candidates for phytoremediation and/or phytomanagement purposes, there is nowadays an urgent need to identify those which can successfully be used under the current scenario of climate change.

The potential indirect effects of climate change on the soil biota present in phytomanaged sites must be also considered, through the abovementioned climate change-induced alterations in plant growth and physiology. Although higher levels of atmospheric CO<sub>2</sub> are a priori not expected to directly affect soil microbial communities (i.e. CO<sub>2</sub> concentrations in the soil are much higher than in the atmosphere), higher atmospheric CO<sub>2</sub> concentrations can indirectly impact on soil microbial communities through higher plant growth, increases in litter deposition and rhizodeposition (often resulting in a stimulation of soil microbial biomass and activity), faster nutrient uptake and water-use efficiency (Phillips et al. 2011; Bardgett et al. 2013; Burns et al. 2013; Alkorta et al. 2017). Such climate change-induced variations of rhizodeposition

patterns, and concomitant changes in the composition and activity of rhizosphere microorganisms, can modify TE bioavailability in soils (Rajkumar et al. 2013), thus potentially affecting plant performance during phytomanagement.

The consequences of climate change (via higher atmospheric CO<sub>2</sub> concentrations, heat waves, extreme droughts, higher temperatures, etc.) on beneficial plant-microorganism interactions (e.g. plant growth-promoting rhizobacteria, endophytes and mycorrhiza) are increasingly being studied (Compant et al. 2005, 2010; Classen et al. 2015; Cavicchioli et al. 2019; Risueño et al. 2020). Plant growth-promoting bacteria and fungi can positively affect water-stressed plants and, then, their inoculation should nowadays be strongly considered for phytomanagement. On the other hand, climate-induced changes in soil temperature and moisture can alter soil processes, such as organic matter decomposition and nutrient cycling (Burns et al. 2013), supported, to a great extent, by the activity of soil microorganisms.

Some phytomanagement practices (e.g. the application of organic amendments, low- or no-tillage practices, grassland implementation and afforestation) have great potential for carbon sequestration and, hence, climate change mitigation. The incorporation of trees in phytomanagement initiatives (e.g. as part of intercropping systems) has also acknowledged positive effects in this respect (Schoeneberger et al. 2012; Alam et al. 2014; Zeng et al. 2019a, b; Brereton et al. 2020).

In theory, a possible option for adaptation to climate change in phytomanagement is to incorporate to the planting scheme as many plant species as possible, and preferably from different vegetation types: grasses, shrubs and trees. Nevertheless, in many situations this is not a realistic, feasible option because the specific plant assemblages established for phytomanagement purposes are determined, to a great extent, on the future land use and on the particular non-food crops intended to be delivered to the local chains processing the harvestable biomass.

Moreover, the conservation of plant biodiversity (e.g. the aforementioned metallophyte diversity) is crucial for adaptation to climate change as part of an “insurance policy”: different species, varieties and ecotypes may be needed in the future as environmental conditions are altered by climate change.

### Promotion of biodiversity under phytomanagement

Under the inherent constraints inevitably derived from the phytomanagement goals, it is certainly possible to promote biodiversity in phytomanaged sites by means of, for instance, growing as many plant species and varieties/ecotypes from different vegetation types (grasses, shrubs and trees) as possible (Table 1). Interestingly, the establishment of different plant species in phytomanaged sites can result in the generation of a wider variety of valuable products and ecosystem services (Evangelou et al. 2015; Pandey and Baudhdh 2018).

Many additional benefits can be obtained when combining different plant species for the phytoremediation and phytomanagement of polluted sites. For instance, the combination of *Pteris vittata* with *Morus alba* and *Broussonetia papyrifera* not only increased the phytoextraction of trace elements but also alleviated phytotoxicity as well (Zeng et al. 2019b). In a similar study, the co-planting of *P. vittata* with *Arundo donax*, *M. alba* and *B. papyrifera* resulted in an improvement of soil health (Zeng et al. 2019a). Furthermore, intercropping with *Paspalum miliaceum* and *Axonopus affinis* was efficient in promoting grapevine growth in Cu-polluted soil by reducing metal bioavailability (De Conti et al. 2019). Interestingly, the combination of tree, shrub and grass species in a metal-polluted soil resulted in a more efficient employment of water resources and a higher biodiversity of soil microorganisms (Parraga-Aguado et al. 2014). In contrast, the co-planting of *Odontarrhena chalcidica* or *Noccaea goesingensis* with *Lotus corniculatus* for Ni removal led to reduced values of shoot biomass (Rosenkranz et al. 2019).

The beneficial effects of co-planting have also been reported for organically polluted and mixed-polluted soils. Wang et al. (2013) reported an enhanced degradation of PAHs, in the presence of trace elements, when *S. alfredii* was combined with *Lolium perenne* or *Ricinus communis*. In agreement with these results, in their studies on intercropping with *Medicago sativa* and *Festuca arundinacea*, Sun et al. (2011) observed higher PAH degradation values under intercropping vs. monoculture. Likewise, intercropping with *M. sativa*, *L. perenne* and *F. arundinacea* improved the degradation of phthalic acid esters. Finally, *F. arundinacea* was also co-planted with *Salix miyabeana* and *M. sativa*, finding out that when crops were cultivated in pairs they showed an enhanced rhizosphere community in terms of the presence of plant growth-promoting bacteria (Brereton et al. 2020).

Aboveground and belowground organisms are closely linked: plants provide organic carbon for soil decomposers and resources for root-associated organisms; in turn, soil decomposers break down dead plant material and regulate plant growth by determining the nutrient supply (Wardle et al. 2004). Different plant species differ in the quantity and quality of litter and root exudates, thus affecting the biomass, activity and diversity (mainly, composition) of soil microbial communities. A more diversified vegetation leads to a higher number of ecological niches and, hence, biodiversity (Risueño et al. 2020). Indeed, higher plant richness results in a higher variety of root exudates and types of litter, thus stimulating biodiversity belowground (Wardle et al. 2004; Haichar et al. 2008). Nonetheless, Li et al. (2015) found no relationship between plant and soil bacterial diversity in an early successional forest, and a negative correlation in a late successional forest. Similarly, Kowalchuk et al. (2000) reported a negative correlation between grassland plant and soil ammonia-oxidising bacterial diversity. These contradictory results point



**Table 1** Ten examples of effects of biodiversity under phytomanagement

Plant species	Contaminants	Main finding	Reference
<i>Pteris vittata</i> co-planted with <i>Morus alba</i> and <i>Broussonetia papyrifera</i>	As, Cd, Pb and Zn	Co-planting alleviated toxicity and improved phytoextraction.	Zeng et al. 2019b
<i>Pteris vittata</i> co-planted with <i>Arundo donax</i> , <i>Morus alba</i> and <i>Broussonetia papyrifera</i>	As, Cd, Pb and Zn	Co-planting enhanced <i>P. vittata</i> growth and metal(oid) accumulation, and improved soil quality.	Zeng et al. 2019a
Co-planting <i>Sedum alfredii</i> with <i>Lolium perenne</i> or <i>Ricinus communis</i>	Metals and PAHs	Co-planting <i>S. alfredii</i> with ryegrass or castor enhanced pyrene and anthracene dissipation.	Wang et al. 2013
Intercropping: <i>Medicago sativa</i> with <i>Festuca arundinacea</i>	PAHs	Removal PAHs under intercropping was higher than under monoculture.	Sun et al. 2011
<i>Odontarrhena chalcidica</i> or <i>Noccaea goesingensis</i> co-planted with <i>Lotus corniculatus</i>	Ni	Intercropping with <i>L. corniculatus</i> tended to decrease the shoot biomass of both species.	Rosenkranz et al. 2019
The grass <i>Piptatherum miliaceum</i> , the shrub <i>Helichrysum decumbens</i> , and the trees <i>Pinus halepensis</i> and <i>Tetraclinis articulata</i>	Metal(loid)s	A diverse set of plant species with contrasting life forms may result in a more efficient employment of water resources and a higher biodiversity not only in relation to flora but also in soil microbes.	Parraga-Aguado et al. 2014
<i>Medicago sativa</i> , <i>Lolium perenne</i> and <i>Festuca arundinacea</i>	Phthalic acid esters (PAEs)	Intercropping with the three species was the most effective treatment for PAEs removal.	Ma et al. 2013
Monocultures and polycultures of <i>Festuca arundinacea</i> , <i>Medicago sativa</i> and <i>Salix miyabeana</i>	Ag, As, Cd, Cr, Cu, Pb, Se and Zn	Co-cropping with the three species was the most robust scenario for remediation of multiple trace element-contaminated soil.	Desjardins et al. 2018
Grapevine was grown in monocropping, intercropping with <i>Paspalum plicatulum</i> and intercropping with <i>Axonopus affinis</i>	Cu	Intercropping with <i>P. plicatulum</i> and <i>A. affinis</i> was efficient in promoting the growth of grapevines at moderate and low levels of Cu contamination by reducing its bioavailability.	De Conti et al. 2019
Co-cropping of <i>Festuca arundinacea</i> , <i>Salix miyabeana</i> and <i>Medicago sativa</i>	Trace elements and persistent organic pollutants (POPs)	The crops cultivated in pairs retained rhizosphere microbiome bacteria involved in plant growth promotion, POP tolerance and degradation, and improved nutrient acquisition	Brereton et al. 2020

out to the vast complexity of the multiple links and interactions between aboveground and belowground diversity (Wardle et al. 2004; De Deyn and Van der Putten 2005; Kardol and Wardle 2010), which, at the moment, are far from being well-understood. Phytomanagement, apart from increasing soil microbial biomass and activity, can induce shifts in the bacterial community structure at both the total community and functional group levels (Touceda-González et al. 2017a). In a study on the effectiveness of dolomite and compost as amendments for enhancing Cu phytostabilisation with *Populus trichocarpa* x *deltoides* cv. Beaupré and *Agrostis gigantea* L., Cu stabilisation and phytomanagement induced positive changes in the microbial community of soil leachates, enriching this community with plant-beneficial bacteria (Giagnoni et al. 2020).

The presence of phytopathogens and root herbivores in the rhizosphere can produce a negative feedback on plant growth, whereas mycorrhizal fungi and plant growth-promoting rhizobacteria can have a positive one on plant growth (Sessitsch et al. 2013; Sura-de Jong et al. 2015). In any case, the evidence for positive or negative links between aboveground and belowground biodiversity is mixed, and not all of the mechanisms by which aboveground organisms affect belowground diversity and vice versa necessarily lead to correlations of species richness in both domains (Hooper et al.

2000). The common perception that belowground biodiversity should follow similar patterns to those of plant diversity during ecosystem development is challenged by Delgado-Baquerizo et al. (2019).

A higher richness of plant species can, for instance, be used to promote the biodegradation of aged polycyclic aromatic compounds in soil: oxygenated PAHs (some of which are more toxic than their related PAHs) can, however, accumulate in soils during such a plant-assisted remediation process (Bandowe et al. 2019).

Through an increase in plant diversity and, hence, in the number of ecological niches and possible habitats, it is also desirable to promote the aboveground and belowground diversity of animals (e.g. arthropods: insects, arachnids, myriapods; earthworms; nematodes; mammals; birds; and so on), of course, always paying close attention to the potential risk of pollutant bioaccumulation and biomagnification (e.g. TE biomagnification along the trophic chain) (Peterson et al. 2003). Interestingly, these animals can act as phytomanagement crop auxiliaries, helping to fight pests, pollinate the cultivated plants, etc. (Verkerk et al. 1998; Ferron and Deguine 2005).

Finally, intercropping systems have been extensively investigated for phytoremediation purposes (Sun et al. 2011; Ma et al. 2013; Wang et al. 2013; Alam et al. 2014; De Conti et al. 2019;

Zeng et al. 2019a, b; Brereton et al. 2020) with additional benefits in terms of aboveground and belowground diversity. Likewise, as individual plant species repeatedly possess a limited range of TE phytoremediation capacities, functional complementarity principles could be of value for the phytoremediation of soils polluted with multiple TEs by means of using assemblages of species (Desjardins et al. 2018).

Biodiversity provides a wide range of values, some of them indirectly such as aesthetic value, cultural value, spiritual value, scientific value and educational value. Arguably, from an anthropocentric point of view, the most important value of biodiversity comes from the ecosystem services it provides. Biodiversity preserves the structure and integrity on which healthy ecosystems depend on to provide the vital ecosystem services on which we rely on.

Among other values of biodiversity, the following two are often discussed when dealing with the conservation of biodiversity and the human use of natural resources: (1) *intrinsic value*: as such, we have the moral responsibility to preserve biodiversity (well-known nature writers such as Henry David Thoreau, John Muir and Aldo Leopold have emphasised the intrinsic value of biodiversity); and (2) *utilitarian value*: as such, focused on the commercial and subsistence benefits (e.g. food, medicines, raw materials, energy) of biodiversity to humankind. Within this utilitarian perspective, the idea is to protect biodiversity so that we can utilise it later for our own benefit. Obviously, this utilitarian value of biodiversity is inextricably linked to the phytomanagement concept. In any case, when designing a phytomanagement initiative, it is unquestionably possible to promote biodiversity within the limits imposed by the specific phytomanagement objectives (e.g. by means of growing as many plant species as possible) with the concomitant potential benefit of obtaining a wider variety of products and ecosystem services.

Anyhow, the biodiversity concept is anything but simple. Among others, it includes the following aspects: *richness* (or the number of species), *evenness* (relative abundances resulting in rare and dominant species), *composition* (in terms of taxonomic groups), *phylogenetic relatedness/distinctiveness* and *spatial and temporal distribution*. Regarding species composition, biological species are certainly not all equal: there are keystone species, foundation species, umbrella species, flagship species, charismatic species, ecosystem engineers, invasive species, indicator species, chemical engineers, biological regulators, etc., leading us to the difficult and arduous challenge to prioritise among them (Vane-Wright et al. 1991).

Some authors proposed to assign more value to those species that lack close relatives, as by maximising the conservation of evolutionary diversity, we maximise genotypic, phenotypic and functional diversity, and, hence, provide ecosystems with the most options to adapt to a changing world (Vane-Wright et al. 1991; Cadotte et al. 2010). Besides, some

species appear to perform phylogenetically narrow processes (e.g. nitrification, atmospheric nitrogen fixation) while others perform phylogenetically broad processes (e.g. denitrification). The former show a lower degree of functional redundancy, compared with the latter.

To assess the influence of phytomanagement practices on biodiversity (such a broad concept) is anything but easy. There are still many unanswered questions that research is yet to answer, e.g. What number of species is a good number? What species composition is best? What degree of phylogenetic distance is more adequate? How differently should we value the different types of species? Are functionally redundant species less valuable than non-functionally redundant species? These questions being answered, we must not take only richness into consideration when promoting biodiversity at polluted sites under phytomanagement. To the best of our expertise and capacities, we must try to consider other relevant aspects also included within the biodiversity concept.

To further complicate matters, biodiversity is difficult to quantify, at least partly, due to the multitude of indices available to measure it (e.g. species richness, Shannon-Wiener entropy, Simpson's index, Berger-Parker index). This is not surprising because of the abovementioned complexity of all the aspects of biodiversity, which inevitably leads to the fact that no single perfect indicator for biodiversity can be devised (Duelli and Obrist 2003). As a matter of fact, the choice of index often depends on the question(s) to be answered, as well as on the specific aspect(s) or entity of biodiversity to be evaluated. Paradoxically, most diversity indices have traditionally relied on three untrue assumptions: (i) all species are equal; (ii) all individuals are equal; and (iii) species abundances have been correctly assessed with appropriate tools and in similar units (Magurran 2004). In any case, although the choice of index(es) depends, to a great extent, on the specific questions and objectives of the study, three of the most commonly used indices are the Margalef's index for richness, the Shannon-Weaver's index for diversity and the Simpson's index for dominance.

Similarly, the use of indices for quantifying functional biodiversity (functional richness, functional evenness and functional divergence) is essential to better understand the links between biodiversity, ecosystem functioning and environmental constraints (Mouchet et al. 2010). Indeed, many studies on the impact of disturbances (e.g. agronomic practices, contamination, climate change, nitrogen deposition) on biological diversity are focused exclusively on structural biodiversity (usually, of only one or a few taxonomic groups). But phytomanagement has a strong functional component related to the provision of ecosystem services. Thus, it is highly beneficial to include both types of biodiversity, i.e. structural and functional diversity, when promoting biodiversity under phytomanagement. Apart from a selection, as wide as possible, of taxonomic groups, an analysis of functional groups,

traits, guilds and so on must be included in phytomanagement initiatives (Kumpiene et al. 2014; Durand et al. 2017; Touceda-González et al. 2017a, b; Xue et al. 2018; Burges et al. 2020).

Although the identification of links between structural and functional biodiversity is undoubtedly a challenging task, such identification is of much value from both an academic/scientific and management point of view. Statistical multivariate analyses, applied to the group of variables used to measure structural and functional diversity, are suitable tools for the establishment of hypotheses regarding the abovementioned links.

The topic of the selection of the best indices to quantify both structural and functional biodiversity is not within the scope of this document. Nonetheless, we encourage the use of various indices for covering as much as possible the different aspects of the term biodiversity: richness, abundance, phylogenetic relatedness, functional traits, etc. Ideally, one should make the best efforts possible to evaluate the effect of phytomanagement practices on the various levels of biodiversity: genetic, species, populations and communities/ecosystems. However, biodiversity is not simply the sum of all ecosystems, species and genetic material, as it represents the variability within and among them.

In particular, for soil microorganisms, the assessment of genetic diversity is indispensable for microbial ecologists since (i) all the current definitions of “species” are inadequate for prokaryotes, among other reasons due to the transfer of genes by horizontal gene transfer; and (ii) most microorganisms cannot be cultivated and so we have no other choice than to study them by means of the application of molecular biology techniques. In consequence, most soil microbial ecologists are nowadays focused on the use of next-generation sequencing techniques (e.g. metabarcoding, metagenomics) for the quantification of soil microbial diversity. But next-generation sequencing has still many technical limitations and then we must be cautious when drawing conclusions about the effect of disturbances or practices (e.g. phytomanagement) on soil microbial diversity.

Most studies on the effect of phytoremediation or phytomanagement practices on microbial diversity are focused on soil microbial communities, especially rhizosphere microorganisms. In this respect, more attention should be paid to plant microbiota and plant microbiomes (e.g. in the phyllosphere) under phytomanagement (Imperato et al. 2019).

Similarly, concerning genetic diversity, there are still many unanswered questions, such as: The more genes the better? Are all genes equally important? Can we talk about “good” genes (e.g. genes involved in contaminant biodegradation pathways) and “bad” genes (e.g. antibiotic resistance genes)? How can be combined data from metagenomic, metatranscriptomic and metaepigenomic studies?

Likewise, when dealing with ecosystem diversity (i.e. the richness and complexity of biological communities, including

trophic levels and ecological processes, together with the chemical and physical environment), additional questions emerge: How many trophic levels do we need? Are all of them equally important? How many species per trophic level are needed?

Regarding the critical links between biodiversity and ecosystem functioning, one should take into consideration the concept of emergent properties, i.e. those new qualities that appear on higher integration levels and represent more than the sum of the low-level components (Reuter et al. 2005). For understanding these emergent properties, the interaction between the different elements must be closely studied (Reuter et al. 2005). In consequence, when possible, key biological interactions should be identified and studied during phytomanagement initiatives, since they support the functioning of the ecosystem and are the basis of emergent properties.

On the other hand, when promoting biodiversity under phytomanagement, it is important to always include organisms from the different levels of the trophic chain. Instead, when evaluating their effect on biodiversity, most phytomanagement initiatives only pay attention to aboveground botanical diversity (richness, composition, vegetation structure) and, occasionally, include some belowground soil biota, in many cases just microorganisms owing to their well-known key role in critical soil processes and, hence, functions and ecosystem services (Xue et al. 2018; Burges et al. 2020). Nonetheless, for a biodiversity assessment with more ecological relevance, it is desirable to include taxonomic groups from the different levels of the trophic chain. As a matter of fact, we should study as many taxonomic groups from the food web as possible (Garrouj et al. 2018; Ali and Khan 2019; Prins et al. 2019).

Simplifying, the aboveground food web includes producers (plants), primary consumers (herbivores) and secondary consumers (predators). Regarding consumers, there is an unresolved debate regarding the benefits and disadvantages associated to the presence of animals in phytomanaged sites. Actually, when dealing with the remediation of polluted sites, in many cases, animals are deliberately excluded, in an attempt to avoid possible ecotoxic effects on exposed animals and, also, to minimise the risk of bioaccumulation and biomagnification (Mann et al. 2011). But animals (e.g. arthropods, earthworms, mammals, birds) can act as phytomanagement crop auxiliaries, helping to fight pests, pollinate the cultivated plants, etc. (Verkerk et al. 1998; Ferron and Deguine 2005).

Pertaining to the soil ecosystem, an amazing diversity of soil organisms make up its food web: bacteria, fungi, algae, protozoa, nematodes, micro-arthropods, earthworms, insects, small vertebrates (mice, moles), etc. Like all food webs, the soil food web is fuelled by primary producers such as plants, lichens, mosses, photosynthetic bacteria (e.g. cyanobacteria) and unicellular algae. The remaining members of the soil biota

obtain energy and carbon by consuming the organic compounds produced by primary producers.

### Adaptive monitoring during phytomanagement

When dealing with long-term monitoring programmes, such as the one for assessing the influence of phytomanagement practices on biodiversity, as time passes, it is inevitable that (i) new analytical techniques, methods and equipment might appear in the market; (ii) different approaches, concepts, ideas, etc. might come up; (iii) changes in the ecosystem developmental stage will occur; (iv) unexpected environmental threats might emerge; and (v) budget fluctuations might threaten the initiative, and so forth (Epelde et al. 2014). For that reason, we propose that the paradigm of adaptive monitoring (this paradigm enables monitoring programmes to evolve iteratively as new information emerges and research questions change) should be incorporated to the long-term monitoring of the effect of phytomanagement practices on biodiversity.

To this purpose, among other aspects, (i) well-formulated, clear and tractable questions must be established at the beginning of the phytomanagement initiative; (ii) a rigorous statistical design must be implemented from the onset of the study, notably accounting for the spatial variability of soil contamination, contaminant exposure and pollutant linkage; and (iii) a conceptual model of the site under phytomanagement must be created (Cundy et al. 2016).

As part of the adaptive monitoring programme, periodically (the time period will depend on the specific phytomanagement initiative), an expert judgement analysis must be organised to revise and, if necessary, update the different aspects that make up the biodiversity monitoring programme. Expert judgement analyses often encourage the forging of partnership between researchers, policy-makers and resource managers, an aspect of the utmost importance in phytomanagement (Cundy et al. 2013).

The accomplishment of economic, social and environmental benefits is a key aspect of phytomanagement (Cundy et al. 2016). In particular, the provisioning of ecosystem services (carbon sequestration, improvement of soil fertility, control of soil erosion, improvement of air quality, climate and water regulation, production of atmospheric oxygen, provision of habitat, etc.) is a crucial component of phytomanagement initiatives (Burges et al. 2018).

The provision of ecosystem services is underpinned by a variety of ecological processes and functions which themselves are driven by biodiversity. Although changes in biodiversity can affect ecosystem processes and, hence, the provision of ecosystem services, in some situations, biomass, species composition, functional traits, etc. are more important than biodiversity itself for the provisioning of those services. Nonetheless, trade-offs between biodiversity and ecosystem services might arise in some situations (Bandowe et al. 2019).

Also, trade-offs and conflicts between the different ecosystem services themselves might also emerge, and, then, it is desirable to select from the onset what specific ecosystem services to promote, and implement measures that minimise conflicts.

### Conclusion

Paraphrasing the three well-known M's of successful trading (Mind, Money management and Method), we can visualise the links between biodiversity and phytomanagement according to three M's of successful phytomanagement: (1) *Mind*: for effective phytomanagement, we must use our mind and creativity to design the best strategy for each specific site and casuistry (here, following the medical aphorism “there are no diseases but sick people”, we can state that “there is no pollution but polluted areas”; all of them are different and require a site-specific assessment). In this respect, biodiversity provides ideas, models and strategies (tested through millions of years of evolution) that we can learn from; (2) *Management*: for successful phytomanagement, we must apply scientifically based adaptive management, especially under the current scenario of climate change. Biodiversity provides a myriad of species, metabolic capabilities, functional traits, etc. which we can use in response to changing conditions; and (3) *Money*: a fruitful phytomanagement will provide economic value through products (crops for biomass-processing technologies) and ecosystem services which can help fuel our bioeconomy. Interestingly, the promotion of biodiversity in phytomanaged sites can result in the generation of a wider variety of valuable products and ecosystem services, while minimising pollutant-induced environmental risks.

**Funding** This work was supported by the Ministry of Economy, Industry and Competitiveness, Spanish Government (AGL2016-76592-R) and European Union within the Interreg SUDOE (PhytoSUDOE-SOE1/P5/E0189).

### References

- Abou-Shanab R, Angle J, Chaney R (2006) Bacterial inoculants affecting nickel uptake by *Alyssum murale* from low, moderate and high Ni soils. *Soil Biol Biochem* 38:2882–2889. <https://doi.org/10.1016/j.soilbio.2006.04.045>
- Agnello AC, Bagard M, van Hullebusch ED, Esposito G, Huguenot D (2016) Comparative bioremediation of heavy metals and petroleum hydrocarbons co-contaminated soil by natural attenuation, phytoremediation, bioaugmentation and bioaugmentation-assisted phytoremediation. *Sci Total Environ* 563–564:693–703. <https://doi.org/10.1016/j.scitotenv.2015.10.061>
- Alam M, Olivier A, Paquette A, Dupras J, Revéret J-P, Messier C (2014) A general framework for the quantification and valuation of ecosystem services of tree-based intercropping systems. *Agrofor Syst* 88: 679–691

- Ali H, Khan E (2019) Trophic transfer, bioaccumulation, and biomagnification of non-essential hazardous heavy metals and metalloids in food chains/webs-concepts and implications for wildlife and human health. *Hum Ecol Risk Assess* 25:1353–1376. <https://doi.org/10.1080/10807039.2018.1469398>
- Alkorta I, Hernández-Allica J, Becerril JM, Amezága I, Albizu I, Garbisu C (2004a) Recent findings on the phytoremediation of soils contaminated with environmentally toxic heavy metals and metalloids such as zinc, cadmium, lead, and arsenic. *Rev Environ Sci Biotechnol* 3: 71–90
- Alkorta I, Hernández-Allica J, Becerril JM, Amezága I, Albizu I, Onaindia M, Garbisu C (2004b) Chelate-enhanced phytoremediation of soils polluted with heavy metals. *Rev Environ Sci Biotechnol* 3:55–70
- Alkorta I, Epelde L, Garbisu C (2017) Environmental parameters altered by climate change affect the activity of soil microorganisms involved in bioremediation. *FEMS Microbiol Lett* 364. <https://doi.org/10.1093/femsle/fix200>
- Anza M, Salazar O, Epelde L, Garbisu C (2018) Data on the selection of biostimulating agents for the bioremediation of soil simultaneously contaminated with lindane and zinc. *Data in Brief* 20:1371–1377. <https://doi.org/10.1016/j.dib.2018.08.203>
- Baker AJM, Ernst WHO, van der Ent A, Malaisse F, Ginocchi R (2010) Metallophytes: the unique biological resource, its ecology and conservation status in Europe, central Africa and Latin America. In: Batty LC, Hallberg KB (eds) *Ecology of industrial pollution*. Cambridge University Press, British Ecological Society, pp 7–40
- Bandowe BAM, Leimer S, Meusel H, Velescu A, Dassen S, Eisenhauer N, Hoffmann T, Oelmann Y, Wilcke W (2019) Plant diversity enhances the natural attenuation of polycyclic aromatic compounds (PAHs and oxygenated PAHs) in grassland soils. *Soil Biol Biochem* 129:60–70. <https://doi.org/10.1016/j.soilbio.2018.10.017>
- Bardgett R, Manning P, Morriën E, De Vries FT (2013) Hierarchical responses of plant-soil interactions to climate change: consequences for the global carbon cycle. *J Ecol* 101:334–343. <https://doi.org/10.1111/1365-2745.12043>
- Barrutia O, Epelde L, García-Plazaola JI, Garbisu C, Becerril JM (2009) Phytoextraction potential of two *Rumex acetosa* L. accessions collected from metalliferous and non-metalliferous sites: effect of fertilization. *Chemosphere* 74:259–264. <https://doi.org/10.1016/j.chemosphere.2008.09.036>
- Barrutia O, Artetxe U, Hernández A, Olano JM, García-Plazaola JI, Garbisu C, Becerril JM (2011a) Native plant communities in an abandoned Pb-Zn mining area of Northern Spain: implications for phytoremediation and germplasm preservation. *Int J Phytorem* 13: 256–270. <https://doi.org/10.1080/15226511003753946>
- Barrutia O, Garbisu C, Epelde L, Sampedro MC, Goicolea MA, Becerril JM (2011b) Plant tolerance to diesel minimizes its impact on soil microbial characteristics during rhizoremediation of diesel-contaminated soils. *Sci Total Environ* 409:4087–4093. <https://doi.org/10.1016/j.scitotenv.2011.06.025>
- Batty LC (2005) The potential importance of mine sites for biodiversity. *Mine Water Environ* 24:101–103
- Becerra-Castro C, Monterroso C, Prieto-Fernández A, Rodríguez-Lamas L, Loureiro-Viñas M, Acea MJ, Kidd PS (2012) Pseudometallophytes colonising Pb/Zn mine tailings: a description of the plant-microorganism-rhizosphere soil system and isolation of metal-tolerant bacteria. *J Hazard Mater* 217–218:350–359. <https://doi.org/10.1016/j.jhazmat.2012.03.039>
- Brereton NJB, Gonzalez E, Desjardins D, Labrecque M, Pitre FE (2020) Co-cropping with three phytoremediation crops influences rhizosphere microbiome community in contaminated soil. *Sci Total Environ* 711:135067. <https://doi.org/10.1016/j.scitotenv.2019.135067>
- Burges A, Epelde L, Benito G, Artetxe U, Becerril JM, Garbisu C (2016) Enhancement of ecosystem services during endophyte-assisted aided phytostabilization of metal contaminated mine soil. *Sci Total Environ* 562:480–492. <https://doi.org/10.1016/j.scitotenv.2016.04.080>
- Burges A, Epelde L, Blanco F, Becerril JM, Garbisu C (2017) Ecosystem services and plant physiological status during endophyte-assisted phytoremediation of metal contaminated soil. *Sci Total Environ* 584:329–338. <https://doi.org/10.1016/j.scitotenv.2016.12.146>
- Burges A, Alkorta I, Epelde L et al (2018) From phytoremediation of soil contaminants to phytomanagement of ecosystem services in metal contaminated sites. *Int J Phytorem* 20:384–397. <https://doi.org/10.1080/15226514.2017.1365340>
- Burges A, Fievet V, Oustrière N, Epelde L, Garbisu C, Becerril JM, Mench M (2020) Long-term phytomanagement with compost and a sunflower - tobacco rotation influences the structural microbial diversity of a Cu-contaminated soil. *Sci Total Environ* 700: 134529. <https://doi.org/10.1016/j.scitotenv.2019.134529>
- Burns RG, DeForest JL, Marxsen J, Sinsabaugh RL, Stromberger ME, Wallenstein MD, Weintraub MN, Zoppini A (2013) Soil enzymes in a changing environment: current knowledge and future directions. *Soil Biol Biochem* 58:216–234. <https://doi.org/10.1016/j.soilbio.2012.11.009>
- Cabello-Conejo MI, Becerra-Castro C, Prieto-Fernández A, Monterroso C, Saavedra-Ferro A, Mench M, Kidd PS (2014) Rhizobacterial inoculants can improve nickel phytoextraction by the hyperaccumulator *Alyssum pintodasilvae*. *Plant Soil* 379:35–50. <https://doi.org/10.1007/s11004-014-2043-7>
- Cadotte MW, Davies TJ, Regetz J, Kembel SW, Cleland E, Oakley TH (2010) Phylogenetic diversity metrics for ecological communities: integrating species richness, abundance and evolutionary history. *Ecol Lett* 13:96–105. <https://doi.org/10.1111/j.1461-0248.2009.01405.x>
- Cardinale BJ, Srivastava DS, Emmett Duffy J, Wright JP, Downing AL, Sankaran M, Jouseau Z (2006) Effects of biodiversity on the functioning of trophic groups and ecosystems. *Nature* 443:989–992
- Cavicchioli R, Ripple WJ, Timmis KN, Azam F, Bakken LR, Baylis M, Behrenfeld MJ, Boetius A, Boyd PW, Classen AT, Crowther TW, Danovaro R, Foreman CM, Huisman J, Hutchins DA, Jansson JK, Karl DM, Koskella B, Mark Welch DB, Martiny JBH, Moran MA, Orphan VJ, Reay DS, Remais JV, Rich VI, Singh BK, Stein LY, Stewart FJ, Sullivan MB, van Oppen MJH, Weaver SC, Webb EA, Webster NS (2019) Scientists' warning to humanity: microorganisms and climate change. *Nat Rev Microbiol* 17:569–586. <https://doi.org/10.1038/s41579-019-0222-5>
- Chan KM, Pringle RM, Ranganathan J, Boggs CL, Chan YL, Ehrlich PR, Haff PK, Heller NE, Al-Khafaji K, Macmynowski DP (2007) When agendas collide: human welfare and biological conservation. *Conserv Biol* 21:51–68. <https://doi.org/10.1111/j.1523-1739.2006.00570.x>
- Chen F, Yang Y, Mi J, Liu R, Hou HP, Zhang SL (2019) Effects of vegetation pattern and spontaneous succession on remediation of potential toxic metal-polluted soil in mine dumps. *Sustainability* 11:397. <https://doi.org/10.3390/su11020397>
- Classen AT, Sundqvist MK, Henning JA, Newman GS, Moore JAM, Cregger MA, Moorhead LC, Courtney M, Patterson CM (2015) Direct and indirect effects of climate change on soil microbial and soil microbial-plant interactions: what lies ahead? *Ecosphere* 6:1–21. <https://doi.org/10.1890/ES15-00217.1>
- Compant S, Duffy B, Nowak J, Clément C, Barka EA (2005) Use of plant growth-promoting bacteria for biocontrol of plant diseases: principles, mechanisms of action, and future prospects. *Appl Environ Microbiol* 71:4951–4959. <https://doi.org/10.1128/AEM.71.9.4951-4959.2005>
- Compant S, Van Der Heijden MGA, Sessitsch A (2010) Climate change effects on beneficial plant-microorganism interactions. *FEMS Microbiol Ecol* 73:197–214. <https://doi.org/10.1111/j.1574-6941.2010.00900>

- Conesa HM, Evangelou MWH, Robinson BH, Schulin R (2012) A critical view of current state of phytotechnologies to remediate soils: still a promising tool? *Sci World J* 2012:Article ID 173829–Article ID 173810. <https://doi.org/10.1100/2012/173829>
- Corzo Remigio A, Chaney RL, Baker AJM, Edraki M, Erskine PD, Echevarria G, van der Ent A (2020) Phytoextraction of high value elements and contaminants from mining and mineral wastes: opportunities and limitations. *Plant Soil* 449:11–37. <https://doi.org/10.1007/s11104-020-04487-3>
- Cundy A, Witter N, Mench M, Friesl W, Müller I, Neu S (2013) Developing principles of sustainability and stakeholder engagement for “gentle” remediation approaches: the European context. *J Environ Manag* 129:283–291. <https://doi.org/10.1016/j.jenvman.2013.07.032>
- Cundy AB, Bardos RP, Puschenreiter M, Mench M, Bert V, Friesl-Hanl W, Müller I, Li XN, Weyens N, Witters N, Vangronsveld J (2016) Brownfields to green fields: realising wider benefits from practical contaminant phytomanagement strategies. *J Environ Manag* 184:67–77. <https://doi.org/10.1016/j.jenvman.2016.03.028>
- De Conti L, Ceretta CA, Melo GWB, Tiecher TL, Silva LOS, Garlet LP, Mimmo T, Cesco S, Brunetto G (2019) Intercropping of young grapevines with native grasses for phytoremediation of Cu-contaminated soils. *Chemosphere* 216:147–156. <https://doi.org/10.1016/j.chemosphere.2018.10.134>
- De Deyn GB, Van der Putten WH (2005) Linking aboveground and belowground diversity. *Trends Ecol Evol* 20:625–633
- Delgado-Baquerizo M, Bardgett RD, Vitousek PM, Maestre FT, Williams MA, Eldridge DJ, Lambers H, Neuhauser S, Gallardo A, García-Velázquez L, Sala OE, Abades SR, Alfaro FD, Berhe AA, Bowker MA, Currier CM, Cutler NA, Hart SC, Hayes PE, Hseu ZY, Kirchmair M, Peña-Ramírez VM, Pérez CA, Reed SC, Santos F, Siebe C, Sullivan BW, Weber-Grullon L, Fierer N (2019) Changes in belowground biodiversity during ecosystem development. *Proc Natl Acad Sci* 116:6891–6896. <https://doi.org/10.1073/pnas.1818400116>
- Dell'Amico E, Cavalca L, Andreoni V (2008) Improvement of *Brassica napus* growth under cadmium stress by cadmium-resistant rhizobacteria. *Soil Biol Biochem* 40:74–84. <https://doi.org/10.1016/j.soilbio.2007.06.024>
- Desjardins D, Brereton NJB, Marchand L, Brisson J, Pitre FE, Labrecque M (2018) Complementarity of three distinctive phytoremediation crops for multiple-trace element contaminated soil. *Sci Total Environ* 610–611:1428–1438. <https://doi.org/10.1016/j.scitotenv.2017.08.196>
- Duelli P, Obrist MK (2003) Biodiversity indicators: the choice of values and measures. *Agric Ecosyst Environ* 98:87–98
- Durand A, Maillard F, Foulon J, Gweon HS, Valot B, Chalot M (2017) Environmental metabarcoding reveals contrasting belowground and aboveground fungal communities from poplar at a Hg phytomanagement site. *Microb Ecol* 74:795–809. <https://doi.org/10.1007/s00248-017-0984-0>
- Dushenkov V, Nanda Kumar PBA, Motto H, Raskin I (1995) Rhizofiltration: the use of plants to remove heavy metals from aqueous streams. *Environ Sci Technol* 29:1239–1245
- Epelde L, Becerril JM, Hernández-Allica J, Barrutia O, Garbisu C (2008) Functional diversity as indicator of the recovery of soil health derived from *Thlaspi caerulescens* growth and metal phytoextraction. *Appl Soil Ecol* 39:299–310. <https://doi.org/10.1016/j.apsoil.2008.01.005>
- Epelde L, Becerril JM, Mijangos I, Garbisu C (2009) Evaluation of the efficiency of a phytostabilization process with biological indicators of soil health. *J Environ Qual* 38:2041–2049. <https://doi.org/10.2134/jeq2009.0006>
- Epelde L, Becerril JM, Kowalchuk GA, Deng Y, Zhou J, Garbisu C (2010) Impact of metal pollution and *Thlaspi caerulescens* growth on soil microbial communities. *Appl Environ Microbiol* 76:7843–7853. <https://doi.org/10.1128/AEM.01045-10>
- Epelde L, Becerril JM, Alkorta I, Garbisu C (2014) Adaptive long-term monitoring of soil health in metal phytostabilization: ecological attributes and ecosystem services based on soil microbial parameters. *Int J Phytorem* 16:971–981. <https://doi.org/10.1080/15226514.2013.810578>
- Evangelou MWH, Papazoglou EG, Robinson BH, Schulin R (2015) Phytomanagement: phytoremediation and the production of biomass for economic revenue on contaminated land. In: Ansari AA, Gill SS, Gill R, Lanza GR, Newman L (eds) *Phytoremediation: management of environmental contaminants*, vol 1. Springer International Publishing, Cham, pp 115–132
- Fässler E, Robinson BH, Stauffer W, Gupta SK, Papritz A, Schulin R (2010) Phytomanagement of metal-contaminated agricultural land using sunflower, maize and tobacco. *Agric Ecosyst Environ* 136:49–58. <https://doi.org/10.1016/j.agee.2009.11.007>
- Ferron P, Deguine J-P (2005) Crop protection, biological control, habitat management and integrated farming. A review. *Agron Sustain Dev* 25:17–24
- Garaiurrebaso O, Garbisu C, Blanco F, Lanzén A, Martín I, Epelde L, Becerril JM, Jechalke S, Smalla K, Grohmann E, Alkorta I (2017) Long-term effects of aided phytostabilization on microbial communities of metal-contaminated mine soil. *FEMS Microbiol Ecol* 93(3). <https://doi.org/10.1093/femsec/fiw252>
- Garbisu C, Alkorta I (2001) Phytoextraction: a cost-effective plant-based technology for the removal of metals from the environment. *Bioresour Technol* 77:229–236
- Garbisu C, Garaiurrebaso O, Epelde L, Grohmann E, Alkorta I (2017) Plasmid-mediated bioaugmentation for the bioremediation of contaminated soils. *Front Microbiol* 8(1966). <https://doi.org/10.3389/fmicb.2017.01966>
- Garrouj M, Marchand L, Frayssinet M, Mench M, Castagneyrol B (2018) Trace element transfer from two contaminated soil series to *Medicago sativa* and one of its herbivores, *Spodoptera exigua*. *Int J Phytorem* 20:650–657. <https://doi.org/10.1080/15226514.2017.1374342>
- Giagnoni L, dos Anjos Borges LG, Giongo A, de Oliveira SA, Ardisson AN, Triplett EW, Mench M, Renella G (2020) Dolomite and compost as amendments for enhancing phytostabilization and increasing microbiota of the leachates from a Cu-contaminated soil. *Agronomy* 10:719. <https://doi.org/10.3390/agronomy10050719>
- Ginocchio R, Baker AJM (2004) Metallophytes in Latin America: a remarkable biological and genetic resource scarcely known and studied in the region. *Rev Chil Hist Nat* 77:185–194. <https://doi.org/10.4067/S0716-078X2004000100014>
- Haichar FZ, Marol C, Berge O, Rangel-Castro JI, Prosser J, Balesdent J, Heulin T, Achouak W (2008) Plant host habitat and root exudates shape soil bacterial community structure. *ISME J* 2:1221–1230
- Haines-Young R, Potschin M (2010) The links between biodiversity, ecosystem services and human well-being. In: Raffaelli DG, Frid CLJ (eds) *Ecosystem ecology: a new synthesis*. Cambridge University Press. British Ecological Society, pp 110–139
- Harrison JG, Griffin EA (2020) The diversity and distribution of endophytes across biomes, plant phylogeny and host tissues: how far have we come and where do we go from here? *Environ Microbiol* 22:2107–2123. <https://doi.org/10.1111/1462-2920.14968>
- Hernández-Allica J, Becerril JM, Zárate O, Garbisu C (2006) Assessment of the efficiency of a metal phytoextraction process with biological indicators of soil health. *Plant Soil* 281:147–158
- Herzig R, Nehnevajova E, Pfister C, Schwytze JP, Ricci A, Keller C (2014) Feasibility of labile Zn phytoextraction using enhanced tobacco and sunflower: results of five- and one-year field-scale experiments in Switzerland. *Int J Phytorem* 16:735–754. <https://doi.org/10.1080/15226514.2013.856846>

- Hooper DU, Bignell DE, Brown VK et al (2000) Interactions between aboveground and belowground biodiversity in terrestrial ecosystems: patterns, mechanisms, and feedbacks. *Bioscience* 50:1049–1061
- Imperato V, Portillo-Estrada M, McAmmond BM, Douwen Y, Van Hamme JD, Gawronski SW, Vangronsveld J, Thijs S (2019) Genomic diversity of two hydrocarbon-degrading and plant growth-promoting *Pseudomonas* species isolated from the oil field of Bóbrka (Poland). *Genes (Basel)* 10:443. <https://doi.org/10.3390/genes10060443>
- IPBES (2019) In: Díaz S, Settele J, Brondizio ES, Ngo HT, Guèze M, Agard J, Arneth A, Balvanera P, Brauman KA, Butchart SHM, Chan KMA, Garibaldi LA, Ichii K, Liu J, Subramanian SM, Midgley GF, Miloslavich P, Molnár Z, Obura D, Pfaff A, Polasky S, Purvis A, Razaque J, Reyers B, Chowdhury RR, Shin YJ, Visseren-Hamakers IJ, Willis KJ, Zayas CN (eds) Summary for policymakers of the global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. IPBES secretariat, Bonn 56 pages
- Jiang C-Y, Sheng X-F, Qian M, Wang Q-Y (2008) Isolation and characterization of a heavy metal-resistant *Burkholderia* sp. from heavy metal-contaminated paddy field soil and its potential in promoting plant growth and heavy metal accumulation in metal-polluted soil. *Chemosphere* 72:157–164. <https://doi.org/10.1016/j.chemosphere.2008.02.006>
- Ju W, Jin X, Liu L, Shen G, Zhao W, Duan C, Fang L (2020) Rhizobacteria inoculation benefits nutrient availability for phytostabilization in copper contaminated soil: drivers from bacterial community structures in rhizosphere. *Appl Soil Ecol Applied* 150:103450. <https://doi.org/10.1016/j.apsoil.2019.103450>
- Kardol P, Wardle DA (2010) How understanding aboveground-belowground linkages can assist restoration ecology. *Trends Ecol Evol* 25:670–679. <https://doi.org/10.1016/j.tree.2010.09.001>
- Kidd P, Mench M, Álvarez-López V et al (2015) Agronomic practices for improving gentle remediation of trace element-contaminated soils. *Int J Phytorem* 17:1005–1037. <https://doi.org/10.1080/15226514.2014.1003788>
- Kolbas A, Kidd P, Guinbertau J, Jaunatre R, Herzig R, Mench M (2015) Endophytic bacteria take the challenge to improve Cu phytoextraction by sunflower. *Environ Sci Pollut Res* 22:5370–5382. <https://doi.org/10.1007/s11356-014-4006-1>
- Kowalchuk GA, Stienstra AW, Stephen JR et al (2000) Changes in the community structure of ammonia oxidizing bacteria during secondary succession of calcareous grasslands. *Environ Microbiol* 2:99–110
- Kumpiene J, Guerri G, Landi L, Pietramellara G, Nannipieri P, Renella G (2009) Microbial biomass, respiration and enzyme activities after *in situ* aided phytostabilization of a Pb- and Cu-contaminated soil. *Ecotoxicol Environ Saf* 72:115–119. <https://doi.org/10.1016/j.ecoenv.2008.07.002>
- Kumpiene J, Bert V, Dimitriou I, Eriksson J, Friesl-Hanl W, Galazka F, Herzig R, Janssen JO, Kidd P, Mench M, Müller I, Neu S, Oustriere N, Puschenreiter M, Renella G, Roumier PH, Siebielec G, Vangronsveld J, Manier N (2014) Selecting chemical and ecotoxicological test batteries for risk assessment of trace element-contaminated soils (phyto)managed by Gentle Remediation Options (GRO). *Sci Total Environ* 496:510–522. <https://doi.org/10.1016/j.scitotenv.2014.06.130>
- Kumpiene J, Renella G, Denys S, Marschner B, Mench M, Adriaensens K, Vangronsveld J, Friesl-Hanl W, Puschenreiter M (2017) Review and evaluation of methods for determining the bioavailable fraction of trace elements in soils. *Pedosphere* 27:389–406. [https://doi.org/10.1016/S1002-0160\(17\)60337-0](https://doi.org/10.1016/S1002-0160(17)60337-0)
- Kuppens T, Van Dael M, Vanreppelen K, Thewys T, Yperman J, Carleer R, Schreurs S, Van Passel S (2015) Techno-economic assessment of fast pyrolysis for the valorization of short rotation coppice cultivated for phytoextraction. *J Clean Prod* 88:336–344. <https://doi.org/10.1016/j.jclepro.2014.07.023>
- Lacalle RG, Gómez-Sagasti MT, Artetxe U, Garbisu C, Becerril JM (2018) *Brassica napus* has a key role in the recovery of the health of soils contaminated with metals and diesel by nano-rhizoremediation. *Sci Total Environ* 618:347–356. <https://doi.org/10.1016/j.scitotenv.2017.10.334>
- Li H, Wang XG, Liang C, Hao Z, Zhou L, Ma S, Li X, Yang S, Yao F, Jiang Y (2015) Aboveground-belowground biodiversity linkages differ in early and late successional temperate forests. *Sci Rep* 5: 12234. <https://doi.org/10.1038/srep12234>
- Ma TT, Teng Y, Luo YM, Christie P (2013) Legume-grass intercropping phytoremediation of phthalic acid esters in soil near an electronic waste recycling site: a field study. *Int J Phytorem* 15:154–167. <https://doi.org/10.1080/15226514.2012.687016>
- Ma Y, Oliveira RS, Nai F, Rajkumar M, Luo Y, Rocha I, Freitas H (2015) The hyperaccumulator *Sedum plumbizincicola* harbors metal-resistant endophytic bacteria that improve its phytoextraction capacity in multi-metal contaminated soil. *J Environ Manag* 156:62–69. <https://doi.org/10.1016/j.jenvman.2015.03.024>
- Madejón E, de Mora AP, Felipe E, Burgos P, Cabrera F (2006) Soil amendments reduce trace element solubility in a contaminated soil and allow regrowth of natural vegetation. *Environ Pollut* 139:40–52. <https://doi.org/10.1016/j.envpol.2005.04.034>
- Magurran AE (2004) Measuring biological diversity, 2nd edn. Blackwell Science Ltd, Oxford
- Malik ZH, Ravindran KC, Sathiyara G (2017) Phytoremediation: a novel strategy and eco-friendly green technology for removal of toxic metals. *Int J Agric Environ Res* 3:1–18
- Mann RM, Vijver MG, Peijnenburg WJGM (2011) Metals and metalloids in terrestrial systems: bioaccumulation, biomagnification and subsequent adverse effects. In: Sánchez-Bayo F, van den Brink PJ, Mann R (eds) Ecological impacts of toxic chemicals. Bentham Science Publishers, pp 43–62
- Massoura ST, Echevarria G, Leclerc-Cessac E, Morel JL (2004) Response of excluder, indicator, and hyperaccumulator plants to nickel availability in soils. *Australian J Soil Res* 42:933–938. <https://doi.org/10.1071/SR03157>
- Mastretta C, Taghavi S, van der Lelie D, Mengoni A, Galardi F, Gonnelli C, Barac T, Boulet J, Weyens N, Vangronsveld J (2009) Endophytic bacteria from seeds of *Nicotiana tabacum* can reduce cadmium phytotoxicity. *Int J Phytoremediat* 11:251–267. <https://doi.org/10.1080/15226510802432678>
- Meglouli H, Fontaine J, Verdin A, Magnin-Robert M, Tisserant B, Hijri M, Lounès-Hadj Saharaoui A (2019) Aided phytoremediation to clean up dioxins/furans-aged contaminated soil: correlation between microbial communities and pollutant dissipation. *Microorganisms* 7: 523. <https://doi.org/10.3390/microorganisms7110523>
- Mench M, Schwitzguébel JP, Schröder P, Bert V, Gawronski S, Gupta S (2009) Assessment of successful experiments and limitations of phytotechnologies: contaminant uptake, detoxification and sequestration, and consequences for food safety. *Environ Sci Pollut Res* 16: 876–900. <https://doi.org/10.1007/s11356-009-0252-z>
- Mench MJ, Dellise M, Bes CM, Marchand L, Kolbas A, le Coustumer P, Oustrière N (2018) Phytomanagement and remediation of Cu-contaminated soils by high yielding crops at a former wood preservation site: sunflower biomass and ionome. *Front Ecol Evol* 6:123. <https://doi.org/10.3389/fevo.2018.00123>
- Mouchet MA, Villéger S, Mason NW et al (2010) Functional diversity measures: an overview of their redundancy and their ability to discriminate community assembly rules. *Funct Ecol* 24:867–876. <https://doi.org/10.1111/j.1365-2435.2010.01695.x>
- Naeem S, Chazdon R, Duffy JE, Prager C, Worm B (2016) Biodiversity and human well-being: an essential link for sustainable

- development. Proc R Soc B 283:20162091. <https://doi.org/10.1098/rspb.2016.2091>
- Olguin EJ, Sanchez-Galvan G, Melo FJ, Hernandez VJ, Gonzalez-Portela RE (2017) Long-term assessment at field scale of floating treatment wetlands for improvement of water quality and provision of ecosystem services in a eutrophic urban pond. Sci Total Environ 584:561–571. <https://doi.org/10.1016/j.scitotenv.2017.01.072>
- Pandey VC, Baudhh K (2018) Phytomanagement of polluted sites - market opportunities in sustainable phytoremediation. Elsevier, p 626. <https://doi.org/10.1016/C2017-0-00586-4>
- Pandey VC, Bajpai O, Singh N (2016) Energy crops in sustainable phytoremediation. Renew Sust Energ Rev 54:58–73. <https://doi.org/10.1016/j.rser.2015.09.078>
- Pardo T, Clemente R, Epelde L, Garbisu C, Bernal MP (2014) Evaluation of the phytostabilisation efficiency in a trace elements contaminated soil using soil health indicators. J Hazard Mater 268:68–76. <https://doi.org/10.1016/j.jhazmat.2014.01.003>
- Parraga-Aguado I, Querejeta JI, Gonzalez-Alcaraz M-N et al (2014) Usefulness of pioneer vegetation for the phytomanagement of metal(loids) enriched tailings: grasses vs. shrubs vs. trees. J Environ Manag 133:51–58. <https://doi.org/10.1016/j.jenvman.2013.12.001>
- Pat-Espadas AM, Loredó Portales R, Amabilis-Sosa LE, Gómez G, Vidal G (2018) Review of constructed wetlands for acid mine drainage treatment. Water 10:1685. <https://doi.org/10.3390/w10111685>
- Paul ALD, Erskine PD, van der Ent A (2018) Metallophytes on Zn-Pb mineralised soils and mining wastes in Broken Hill, NSW, Australia. Australian J Bot 66:124–133. <https://doi.org/10.1071/BT17143>
- Peterson LR, Trivett V, Baker AJM, Aguiar C, Pollard AJ (2003) Spread of metals through an invertebrate food chain as influenced by a plant that hyperaccumulates nickel. Chemoecology 13:103–108. <https://doi.org/10.1007/s00049-003-0234-4>
- Phillips R, Finzi A, Bernhardt E (2011) Enhanced root exudation induces microbial feedbacks to N cycling in a pine forest under long-term CO<sub>2</sub> fumigation. Ecol Lett 14:187–194. <https://doi.org/10.1111/j.1461-0248.2010.01570.x>
- Prins CN, Hantzis LJ, Valdez-Barillas JR, Cappa JJ, Fakra SC, Milano de Tomasel C, Wall DH, Pilon-Smits EAH (2019) Getting to the root of selenium hyperaccumulation – localization and speciation of root selenium and its effects on nematodes. Soil Systems 3:47. <https://doi.org/10.3390/soilsystems3030047>
- Pulighe G, Bonati G, Colangeli M, Morese MM, Traverso L, Lupia F, Khawaja C, Janssen R, Fava F (2019) Ongoing and emerging issues for sustainable bioenergy production on marginal lands in the Mediterranean regions. Renew Sust Energ Rev 103:58–70. <https://doi.org/10.1016/j.rser.2018.12.043>
- Rajkumar M, Prasad MNV, Swaminathan S, Freitas H (2013) Climate change driven plant-metal-microbe interactions. Environ Int 53:74–86. <https://doi.org/10.1016/j.envint.2012.12.009>
- Reeves RD, Baker AJM, Jaffré T, Erskine PD, Echevarria G, van der Ent A (2017a) A global database for plants that hyperaccumulate metal and metalloid trace elements. New Phytol 218:407–411. <https://doi.org/10.1111/nph.14907>
- Reeves RD, Baker AJM, Jaffre T, Erskine PD, Echevarria G, van der Ent A (2017b) A global database for plants that hyperaccumulate metal and metalloid trace element. New Phytol 218:407–411. <https://doi.org/10.1111/nph.14907>
- Reuter H, Hölker F, Middelhoff U, Jopp F, Eschenbach C, Breckling B (2005) The concepts of emergent and collective properties in individual-based models - summary and outlook of the Bornhöved case studies. Ecol Model 186:489–501. <https://doi.org/10.1016/j.ecolmodel.2005.02.014>
- Risueño Y, Petri C, Conesa HM (2020) The importance of edaphic niches functionality for the sustainability of phytomanagement in semiarid mining impacted ecosystems. J Environ Manag 266:110613. <https://doi.org/10.1016/j.jenvman.2020.110613>
- Robinson BH, McIvor I (2013) Phytomanagement of contaminated sites using poplars and willows. In: Leung DWM (ed) Recent advances towards improved phytoremediation of heavy metal pollution. Bentham Science, Arab Emirates, pp 119–133
- Robinson BH, Banuelos G, Conesa HM, Evangelou MWH, Schulin R (2009) The phytomanagement of trace elements in soil. Crit Rev Plant Sci 28:240–266. <https://doi.org/10.1080/07352680903035424>
- Rosenkranz T, Hipfinger C, Ridard C, Puschenreiter M (2019) A nickel phytomining field trial using *Odontarrhena chalcidica* and *Noccaea goesingensis* on an Austrian serpentine soil. J Environ Manag 242: 522–528. <https://doi.org/10.1016/j.jenvman.2019.04.073>
- Sandifer PA, Sutton-Grier AE, Ward BP (2015) Exploring connections among nature, biodiversity, ecosystem services, and human health and well-being: opportunities to enhance health and biodiversity conservation. Ecosystem Services 12:1–15. <https://doi.org/10.1016/j.ecoser.2014.12.007>
- Schoeneberger M, Bentrup G, de Gooijer H, Soolanayakanahally R, Sauer T, Brandle J, Zhou X, Current D (2012) Branching out: agroforestry as a climate change mitigation and adaptation tool for agriculture. J Soil Water Conserv 67:128A–136A. <https://doi.org/10.2489/jswc.67.5.128A>
- Schröder P, Navarro-Aviñó J, Azaizeh H, Goldhirsh AG, DiGregorio S, Komives T, Langergraber G, Lenz A, Maestri E, Memon AR, Ranalli A, Sebastiani L, Smrcek S, Vanek T, Vuilleumier S, Wissing F (2007) Using phytoremediation technologies to upgrade waste water treatment in Europe. Environ Sci Pollut Res 14:490–497
- Séleck M, Bizoux JP, Colinet G, Faucon MP, Guillaume A, Meerts P, Piqueray J, Mahy G (2013) Chemical soil factors influencing plant assemblages along copper-cobalt gradients: implications for conservation and restoration. Plant Soil 373:455–469
- Sessitsch A, Kuffner M, Kidd P, Vangronsveld J, Wenzel WW, Fallmann K, Puschenreiter M (2013) The role of plant-associated bacteria in the mobilization and phytoextraction of trace elements in contaminated soils. Soil Biol Biochem 60:182–194. <https://doi.org/10.1016/j.soilbio.2013.01.012>
- Simek M, Elhottova D, Mench M et al (2017) Greenhouse gas emissions from a Cu-contaminated soil remediated by *in situ* stabilization and phytomanaged by a mixed stand of poplar, willows, and false indigo-bush. Int J Phytorem 19:976–984. <https://doi.org/10.1080/15226514.2016.1267706>
- Sun M, Fu D, Teng Y, Shen Y, Luo Y, Li Z, Christie P (2011) *In situ* phytoremediation of PAH-contaminated soil by intercropping alfalfa (*Medicago sativa* L.) with tall fescue (*Festuca arundinacea* Schreb.) and associated soil microbial activity. J Soils Sediments 11:980–989. <https://doi.org/10.1007/s11368-011-0382>
- Sura-de Jong M, Reynolds RJ, Richterova K, Musilova L, Hrochova I, Frantik T, Sakmaryova I, Strejcek M, Cochran A, Staicu L, Cappa JJ, van der Lelie D, Pilon-Smits EAH (2015) Selenium hyperaccumulators harbor a diverse endophytic bacterial community characterized by high selenium resistance and plant growth promoting properties. Front Plant Sci 6:113. <https://doi.org/10.3389/fpls.2015.00113>
- Thewys T, Witters N, Meers E, Vangronsveld J (2010) Economic viability of phytoremediation of a cadmium contaminated agricultural area using energy maize. Part II: economics of anaerobic digestion of metal contaminated maize in Belgium. Int J Phytorem 12:663–679. <https://doi.org/10.1080/15226514.2010.493188>
- Touceda-González M, Prieto-Fernández A, Renella G, Giagnoni L, Sessitsch A, Brader G, Kumpiene J, Dimitriou I, Eriksson J, Friesl-Hanl W, Galazka R, Janssen J, Mench M, Müller I, Neu S, Puschenreiter M, Siebielec G, Vangronsveld J, Kidd PS (2017a) Microbial community structure and activity in trace element contaminated soils phytomanaged by Gentle Remediation Options (GRO). Environ Pollut 231:237–251. <https://doi.org/10.1016/j.envpol.2017.07.097>



- Touceda-González M, Álvarez-López V, Prieto-Fernández A, Rodríguez-Garrido B, Trasar-Cepeda C, Mench M, Puschenreiter M, Quintela-Sabaris C, Macías-García F, Kidd PS (2017b) Aided phytostabilisation reduces metal toxicity, improves soil fertility and enhances microbial activity in Cu-rich mine tailings. *J Environ Manag* 186:301–313. <https://doi.org/10.1016/j.jenvman.2016.09.019>
- Truyens S, Jambon I, Croes S, Janssen J, Weyens N, Mench M, Carleer R, Cuypers A, Vangronsveld J (2014) The effect of long-term Cd and Ni exposure on seed endophytes of *Agrostis capillaris* and their potential application in phytoremediation of metal-contaminated soils. *Int J Phytorem* 16:643–659. <https://doi.org/10.1080/15226514.2013.837027>
- van der Ent A, Baker AJM, Reeves RD, Pollard AJ, Henk Schat H (2013) Hyperaccumulators of metal and metalloid trace elements: facts and fiction. *Plant Soil* 362:319–334. <https://doi.org/10.1007/s11104-012-1287>
- van der Ent A, Baker AJM, Reeves RD, Pollard AJ, Schat H (2015a) A commentary on “Toward a more physiologically and evolutionarily relevant definition of metal hyperaccumulation in plants”. *Front Plant Sci* 6:1–3. <https://doi.org/10.3389/fpls.2015.00554>
- van der Ent A, Baker AJM, Reeves RD, Chaney RL, Anderson CWN, Meech JA, Erskine PD, Simonnot M-O, Vaughan J, Morel JL, Echevarria G, Fogliani B, Rongliang Q, Mulligan DR (2015b) Agromining: farming for metals in the future? *Environ Sci Technol* 49:4773–4780. <https://doi.org/10.1021/es506031u>
- Vane-Wright RI, Humphries CJ, Williams PH (1991) What to protect - systematics and the agony of choice. *Biol Conserv* 55:235–254
- Vangronsveld J, Herzig R, Weyens N, Boulet J, Adriaensens K, Ruttens A, Thewys T, Vassilev A, Meers E, Nehnevajova E, van der Lelie D, Mench M (2009) Phytoremediation of contaminated soils and groundwater: lessons from the field. *Environ Sci Pollut Res Int* 16:765–794. <https://doi.org/10.1007/s11356-009-0213-6>
- Verkerk RHJ, Leather SR, Wright DJ (1998) The potential for manipulating crop-pest-natural enemy interactions for improved insect pest management. *Bull Entomol Res* 88:493–501
- Villaverde J, Láiz L, Lara-Moreno A, González-Pimentel JL, Morillo E (2019) Bioaugmentation of PAH-contaminated soils with novel specific degrader strains isolated from a contaminated industrial site. Effect of hydroxypropyl- $\beta$ -cyclodextrin as PAH bioavailability enhancer. *Front Microbiol* 14 November 2019. <https://doi.org/10.3389/fmicb.2019.02588>
- Vincent Q, Auclerc A, Beguiristain T, Leyval C (2018) Assessment of derelict soil quality: abiotic, biotic and functional approaches. *Sci Total Environ* 614-614:990–1002. <https://doi.org/10.1016/j.scitotenv.2017.09.118>
- Wang K, Huang H, Zhu Z, Li T, He Z, Yang X, Alva A (2013) Phytoextraction of metals and rhizoremediation of PAHs in co-contaminated soil by co-planting of *Sedum alfredii* with ryegrass or castor. *Int J Phytorem* 15:283–298. <https://doi.org/10.1080/15226514.2012.694501>
- Wardle DA, Bardgett RD, Klironomos JN, Setälä H, van der Putten W, Wall DH (2004) Ecological linkages between aboveground and belowground biota. *Science* 304:1629–1633
- Weyens N, Truyens S, Saenen E, Boulet J, Dupae J, Taghavi S, Lelie D, Carleer R, Vangronsveld J (2011) Endophytes and their potential to deal with co-contamination of organic contaminants (toluene) and toxic metals (nickel) during phytoremediation. *Int J Phytoremediation* 13:244–255. <https://doi.org/10.1080/15226511003753920>
- Whiting SN, Reeves RD, Baker AJM (2002) Mining, metallophytes and land reclamation. *Min Environ Manag* 10:11–16
- Whiting SN, Reeves RD, Richards DG, Johnson MS, Cooke JA, Malaisse F, Paton A, Smith JAC, Angle JS, Chaney RL, Ginocchio R, Jaffre T, Johns R, McIntyre T, Purvis OW, Salt DE, Schat H, Zhao FJ, Baker AJM (2004) Research priorities for conservation of metallophyte biodiversity and their potential for restoration and site remediation. *Restor Ecol* 12:106–116. <https://doi.org/10.1111/j.1061-2971.2004.00367.x>
- Xue K, Zhou J, Van Nostrand JD, Mench M, Bes C, Giagnoni L, Arenella M, Renella G (2018) Functional activity and functional gene diversity of a Cu-contaminated soil remediated by aided phytostabilization using compost, dolomitic limestone and a mixed tree stand. *Environ Pollut* 242:229–238. <https://doi.org/10.1016/j.envpol.2018.06.057>
- Yang W, Zhao F, Wang Y, Ding Z, Yang X, Zhu Z (2020a) Differences in uptake and accumulation of copper and zinc by *Salix* clones under flooded versus non-flooded conditions. *Chemosphere* 241:125059. <https://doi.org/10.1016/j.chemosphere.2019.125059>
- Yang C, Ho Y-H, Makita R, Inoue C, Chien M-F (2020b) A multifunctional rhizobacterial strain with wide application in different ferns facilitates arsenic phytoremediation. *Sci Total Environ* 712:134504. <https://doi.org/10.1016/j.scitotenv.2019.134504>
- Zaidi S, Usmani S, Singh BR, Musarrat J (2006) Significance of *Bacillus subtilis* strain SJ-101 as a bioinoculant for concurrent plant growth promotion and nickel accumulation in *Brassica juncea*. *Chemosphere* 64:991–997. <https://doi.org/10.1016/j.chemosphere.2005.12.057>
- Zeng P, Guo Z, Xiao X, Peng C, Huang B, Feng WL (2019a) Complementarity of co-planting a hyperaccumulator with three metal(loid)-tolerant species for metal(loid)-contaminated soil remediation. *Ecotoxicol Environ Saf* 169:306–315. <https://doi.org/10.1016/j.ecoenv.2018.11.017>
- Zeng P, Guo ZH, Xiao XY, Peng C, Feng WL, Xin LQ, Xu Z (2019b) Phytoextraction potential of *Pteris vittata* L. co-planted with woody species for As, Cd, Pb and Zn in contaminated soil. *Sci Total Environ* 650:594–603. <https://doi.org/10.1016/j.scitotenv.2018.09.055>
- Zhang XB, Liu P, Yang YS, Chen WR (2007) Phytoremediation of urban wastewater by model wetlands with ornamental hydrophytes. *J Environ Sci (China)* 19:902–909. [https://doi.org/10.1016/S1001-0742\(07\)60150-8](https://doi.org/10.1016/S1001-0742(07)60150-8)

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.